

Ca²⁺-INDUCED COLD-SET GELATION OF WHEY PROTEIN ISOLATE FIBRILS

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ABSTRACT:

In this paper we describe the rheological behaviour of Ca²⁺-induced cold-set gels of whey protein mixtures. Cold-set gels are important applications for products with a low thermal stability. In previous work [1], we determined the state diagram for whey protein mixtures that were heated for 10 h at pH 2 at 80°C. Under these conditions, the major whey protein, β -lactoglobulin (β -lg), forms fibrils. When whey protein mixtures are heated at protein concentrations in the liquid solution regime of the state diagram, cold-set gels can be formed by adding Ca²⁺ ions at pH 7. We studied the rheological behaviour of cold-set gels for various sample compositions for whey protein mixtures. When keeping the total whey protein concentration constant, the elastic modulus, G' , for the cold-set gels decreased for increasing α -lactalbumin and bovine serum albumin ratios, because less material (β -lg fibrils) was available to form a gel network. In the cold-set gels the interactions between the β -lg fibrils induced by the calcium ions are dominant. The β -lg fibrils are forming the cold-set gel network and therefore determine the gel strength. α -Lactalbumin and bovine serum albumin are not incorporated in the stress-bearing structure of the gels.

ZUSAMMENFASSUNG:

In diesem Artikel wird das rheologische Verhalten von Ca²⁺-induzierten, kaltgehärteten Gelen aus Mischungen von Molke-Protein beschrieben. Kaltgehärtete Gele sind wichtige Anwendungen für Produkte mit einer geringen thermischen Stabilität. Schon in früheren Arbeiten wurde von uns das Zustandsdiagramm von Molke-Protein-Mischungen, die für 10 Stunden bei einem pH 2 auf 80°C erhitzt wurden, erstellt [1]. Unter diesen Bedingungen formt das häufigste und daher wichtigste Molke-Protein, β -Lactoglobulin (β -lg), Fibrillen. Die flüssige Phase des Zustandsdiagramms von Molke-Protein kann, bei einem pH-Wert von 7, durch Erhitzen und Zugabe von Ca²⁺-Ionen kaltgehärtete Gele bilden. Dies wird für verschiedene Mischungen von Molke-Proteinen untersucht. Bei gleich bleibender totaler Proteinkonzentration und bei steigendem α -Lactalbumin Gehalt und Bovin Serum Albumin Gehalt wird der elastische Modul für kaltgehärtete-Gele kleiner, da weniger Material (β -Lactoglobulin Fibrillen) zur Formung eines Gelnetzwerkes vorhanden ist. In kaltgehärteten Gelen sind die durch Kalziumionen induzierten Wechselwirkungen zwischen den β -lg Fibrillen dominant. Die β -lg Fibrillen formen das kaltgehärtete Gel-Netzwerk und bestimmen daher die Gelstärke. α -Lactalbumin und BSA wurden nicht in diese Strukturen eingebaut.

RÉSUMÉ:

Dans cet article, nous décrivons le comportement rhéologique de gels formés à partir de mixtures de protéines du petit lait, et induits à froid par l'addition de Ca²⁺. Les gels formés à froid sont utilisés de façon importante dans les produits montrant une faible stabilité thermique. Dans un travail précédent [1], nous avons déterminé le diagramme de phase pour les mixtures de petit lait qui ont été chauffées durant 10 heures à 80°C et à pH 2. Sous ces conditions, la protéine majoritaire, la β -lactoglobuline (β -lg), forme des fibrilles. Quand les mixtures sont chauffées avec des concentrations en protéines correspondant au régime de solution liquide du diagramme de phase, des gels peuvent être formés à froid par addition d'ions Ca²⁺ à pH 7. Nous avons étudié le comportement rhéologique des gels formés à froid pour des échantillons comprenant des compositions variées de mixtures de protéines. Lorsque la concentration totale en protéines est maintenue constante, le module élastique G' des gels formés à froid décroît avec l'augmentation des ratios de α -lactalbumine et de d'albumine de sérum bovin, parce que moins de matériau (fibrilles de β -lg) est disponible pour former un réseau de gel. Dans les gels formés à froid, les interactions entre les fibrilles de β -lg forment le réseau du gel et donc déterminent la rigidité du gel. L' α -lactalbumine et l'albumine de sérum bovin ne sont pas incorporées dans la structure du gel qui supporte la contrainte.

KEY WORDS: cold-set gelation, whey protein isolate, β -lactoglobulin, fibrils

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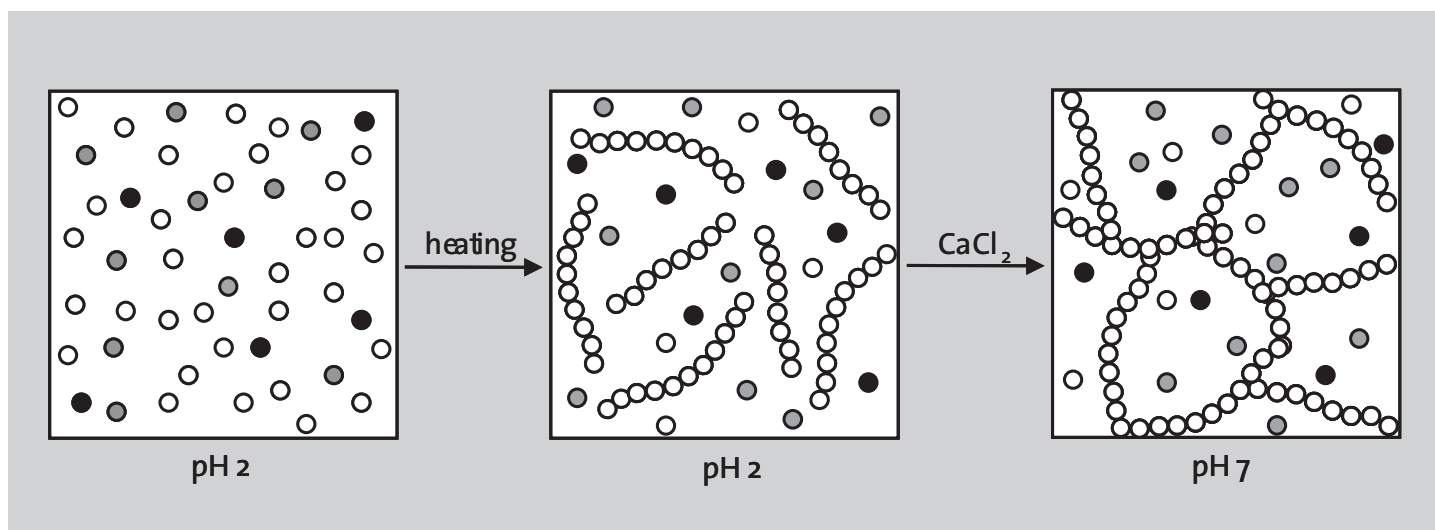
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ples (see the right vertical planes in Figs. 3a and 3c). Therefore we can conclude that β -lg is dominant, and that α -lac and BSA do not contribute to the cold-set gel network. Under the conditions used in our experiments, α -lac and BSA do not form fibrils and form at most small aggregates [1], whereas β -lg forms long fibrils upon heating at pH 2. In heated whey protein samples depletion interactions between the long β -lg fibrils can be induced by monomers or small aggregates. This can result in attractive interactions between β -lg fibrils, causing a decrease in the minimum gelation concentration of β -lg for heat-set gels at pH 2 [1]. The small α -lac and BSA aggregates apparently are too small to contribute to the Ca^{2+} -induced cold-set gels at pH 7. The β -lg fibrils form the Ca^{2+} -induced cold-set gel network and therefore determine the gel strength. The attractive depletion interactions that play a role in the heat-set gelation at pH 2 do not contribute as much in the cold-set gels as in the heat-set pH 2 gels. In the cold-set gels the interactions between the β -lg fibrils, induced by the calcium ions, are dominant. Therefore a lower G' is found when the α -lac or BSA concentration in the samples increases, and the total protein concentration is kept constant, because less material (β -lg fibrils) is available to form a gel network. A schematic representation of the Ca^{2+} -induced cold-set gelation method for a mixture of whey proteins is shown in Fig. 5.

From the values of G' obtained from the various concentration curves, the critical percolation concentration, c_p , and the scaling component, t , were calculated. Here we define the minimal gelation concentration, based on the total protein concentration, as the critical percolation concentration, c_p . The scaling relation that we used is $G' \sim (c - c_p)^t$, where c is the concentration of monomers. Both c_p and t were determined using the method described by Van der Linden and Sagis [25]. This method is a graphical method that uses plots of

$(G')^{1/t}$ versus c and extrapolates these plots to $(G')^{1/t} = 0$. In this procedure we make use of the fact that independent of the value of t the curves must all intersect the concentration axis at the same value. When the assumed value for t is close to the actual value, the plot will be linear. If t is too small or too large, the lines are curved. From the plots of $(G')^{1/t}$ versus c for various t , we selected those t values that give an approximately straight line. From these plots, an average value of c_p was determined. We plotted $\log G'$ versus $\log (c - c_p)$, using the different values for c_p obtained from the estimated t values. For each of the values for c_p , we determined a new value for t and averaged these values. In Tab. 2 the calculated values for c_p and t are shown. The c_p values are all in a narrow range between 0.45 and 0.94 and do not show a systematic dependence on composition of the samples. This confirms that α -lac and BSA do not contribute to the gel network. For almost all samples the calculated values for t were between 1.8 and 2.0, which indicates isotropic force percolation. Isotropic force percolation assumes that nearest neighbours in the network interact through an isotropic force.

4 CONCLUSION

For the Ca^{2+} -induced cold-set gelation, performed with whey protein samples in the liquid solution regime of the state diagram, the minimal gelation concentration, c_p , was not affected by the addition of α -lac or BSA to β -lg (within the accuracy of the experiments). However, the G' for cold-set gels at equal total protein concentra-

Sample	c_p [wt%] total protein concentration	t [-]
β -lg	0.57 ± 0.04	2.24 ± 0.12
β -lg/ α -lac = 90/10	0.48 ± 0.07	2.18 ± 0.18
β -lg/ α -lac = 80/20	0.45 ± 0.08	1.94 ± 0.21
β -lg/BSA = 90/10	0.53 ± 0.07	1.97 ± 0.15
β -lg/BSA = 83/17	0.50 ± 0.04	1.75 ± 0.12
β -lg/BSA = 70/30	0.94 ± 0.04	1.80 ± 0.11
WPI	0.75 ± 0.15	1.97 ± 0.44
purified WPI	0.58 ± 0.12	1.91 ± 0.33

Samples were first heated at 2.5 wt % for 10 h at pH 2 and 80°C, subsequently cooled to room temperature. The values for G' obtained from Ca^{2+} -induced cold gelation with the addition of 0.01 M CaCl_2 at pH 7 was measured at 25°C after 3 h of gelation.

Figure 5: Schematic representation of the Ca^{2+} -induced cold-set gelation method for a mixture of whey proteins; (○) β -lg; (●) α -lac; (●) BSA. First we have whey proteins in solution at pH 2. This solution is heated at a temperature above the denaturation temperature of the protein, and at a total protein concentration below the heat-induced gelation concentration. Under the conditions used β -lg fibrils are formed during heating. The fibril solution is cooled and the pH is adjusted from pH 2 to pH 7. Gelation is induced by addition of CaCl_2 .

Table 2: Calculated values for c_p and t for the total whey protein concentrations.

tions was found to decrease for increasing α -lac or BSA concentrations, because less material (β -lg fibrils) was available to form a gel network. The attractive depletion interactions that appear to play a role in the heat-set gelation at pH 2 do not contribute as much in the cold-set gels. In the cold-set gels the interactions between the β -lg fibrils, induced by the calcium ions, are dominant. The β -lg fibrils are forming the cold-set gel network and therefore determine the gel strength.

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REFERENCES

- [1] Bolder SG, Hendrickx H, et al.: Fibril assemblies in aqueous whey protein mixtures, *J. Agric. Food Chem.* 54 (2006) 4229-4234.
- [2] Aymard P, Nicolai T, et al.: Static and dynamic scattering of β -lactoglobulin aggregates formed after heat-induced denaturation at pH 2, *Macromolecules* 32 (1999) 2542-2552.
- [3] Verheul M: Aggregation and gelation of whey proteins, Universiteit Twente, The Netherlands (1997) pp.153.
- [4] Bryant CM, McClements DJ: Molecular basis of protein functionality with special consideration of cold-set gels derived from heat-denatured whey, *Trends Food Sci. Technol.* 9 (1998) 143-151.
- [5] Doi E: Gels and gelling of globular proteins, *Trends Food Sci. Technol.* 4 (1993) 1-5.
- [6] Aymard P, Durand D, et al.: The effect of temperature and ionic strength on the dimerisation of β -lactoglobulin, *Int. J. Biol. Macromol.* 19 (1996) 213-221.
- [7] Gimel JC, Durand D, et al.: Structure and distribution of aggregates formed after heat-induced denaturation of globular proteins., *Macromolecules* 27 (1994) 583-589.
- [8] Harwalkar VR, Kalab M: Thermal denaturation and aggregation of β -lactoglobulin in solution. Electron microscopic study, *Milchwiss. - Milk Sci. Int.* 40 (1985) 65-68.
- [9] Langton M, Hermansson A-M: Fine-stranded and particulate gels of β -lactoglobulin and whey protein at varying pH, *Food Hydrocolloids* 5 (1992) 523-539.
- [10] Le Bon C, Nicolai T, et al.: Growth and structure of aggregates of heat-denatured β -lactoglobulin, *Int. J. Food Sci. Technol.* 34 (1999)
- [11] Schokker EP, Singh H, et al.: Characterization of intermediates formed during heat-induced aggregation of β -lactoglobulin AB at neutral pH, *Int. Dairy J.* 9 (1999) 791-800.
- [12] Stading M, Langton M, et al.: Inhomogeneous fine-stranded β -lactoglobulin gels., *Food Hydrocolloids* 6 (1992) 455-470.
- [13] Arnaudov LN, de Vries R, et al.: Multiple steps during the formation of β -lactoglobulin fibrils, *Bio-macromolecules* 4 (2003) 1614-1622.
- [14] Ikeda S, Morris VJ: Fine-stranded and particulate aggregates of heat-denatured whey proteins visualized by atomic force microscopy, *Bio-macromolecules* 3 (2002) 382-389.
- [15] Kavanagh GM, Clark AH, et al.: Heat-induced gelation of globular proteins: part 3. Molecular studies on low pH β -lactoglobulin gels, *Int. J. Biol. Macromol.* 28 (2000) 41-50.
- [16] Renard D, Axelos MAV, et al.: Investigation of sol-gel transitions of β -lactoglobulin by rheological and small-angle neutron scattering measurements, *Food Macromolecules and Colloids*, Dickinson E and Lorient Ds (Eds.), Royal Society of Chemistry: Cambridge, UK (1995) 391-399.
- [17] Veerman C, Ruis H, et al.: Effect of electrostatic interactions on the percolation concentration of fibrillar β -lactoglobulin gels, *Biomacromolecules* 3 (2002) 869-873.
- [18] Veerman C, Baptist H, et al.: A new multi-step Ca^{2+} -induced cold gelation process for β -lactoglobulin, *J. Agric. Food Chem.* 51 (2003) 3880-3885.
- [19] Bryant CM, McClements DJ: Optimizing preparation conditions for heat-denatured whey protein solutions to be used as cold-gelling ingredients, *J. Food Sci.* 65 (2000) 259-263.
- [20] Bryant CM, McClements DJ: Influence of NaCl and CaCl_2 on cold-set gelation of heat-denatured whey protein, *J. Food Sci.* 65 (2000) 801-804.
- [21] Hongsprabhas P, Barbut S: Ca^{2+} -induced gelation of whey protein isolate: effects of pre-heating, *Food Res. Int.* 29 (1996) 135-139.
- [22] Hongsprabhas P, Barbut S: Structure-forming processes in Ca^{2+} -induced whey protein isolate cold gelation, *Int. Dairy J.* 7 (1997) 827-834.
- [23] Hongsprabhas P, Barbut S, et al.: The structure of cold-set whey protein isolate gels with Ca^{2+} , *Lebensm. Wiss. Technol. - Food Sci. Technol.* 32 (1999) 196-202.
- [24] Weinbreck F, de Vries R, et al.: Complex coacervation of whey proteins and gum arabic, *Bio-macromolecules* 4 (2003) 293-303.
- [25] van der Linden E, Sagis LMC: Isotropic force percolation in protein gels, *Langmuir* 17 (2001) 5821-5824.

